

Instructions for use

FlexStar[®]

CT & NG PCR Detection Mix 1.5

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FlexStar[®]

CT & NG PCR Detection

Mix 1.5

For research use only!

(RUO)



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1. Application

The FlexStar® CT & NG PCR Detection Mix 1.5 is a reagent system, based on real-time PCR technology, for the qualitative detection and differentiation of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* specific DNA.

For research use only (RUO)! Not for use in diagnostic procedures.

2. Product content

The FlexStar® CT & NG PCR Detection Mix 1.5 contains the following components:

Table 1: Kit components

Lid color	Component	Number of tubes	Nominal volume [µl/tube]
Blue	Detection Mix ¹⁾	8	60
Red	PC ²⁾	2	250
White	NTC ³⁾	2	250

¹⁾ Contains biological material of animal origin

²⁾ Positive Control (CT and NG specific DNA)

³⁾ No Template Control (negative control)

3. Storage

- The FlexStar® CT & NG PCR Detection Mix 1.5 is shipped on dry ice. The product components should arrive frozen. If one or more components are not frozen upon receipt, or if tubes have been compromised during shipment, contact Altona Diagnostics technical support for assistance (see chapter 9. Technical assistance).
- All components should be stored at -25 °C to -15 °C upon arrival.
- Repeated thawing and freezing of the Detection Mix component should be avoided, as this might affect the performance of the product.
- Repeated thawing and freezing of the Positive Control (PC) and No Template Control (NTC) (more than 4 times) should be avoided, as this might affect the performance of the product.
- Storage at room temperature (max. +30 °C) should not exceed a period of 2 hours.
- Protect the Detection Mix component from light.

4. Product description

The FlexStar® CT & NG PCR Detection Mix 1.5 is a reagent system. Used in combination with the FlexStar® (RT-)PCR Amplification Mix 1.5 it allows the qualitative detection and differentiation of *Chlamydia trachomatis* (CT) and *Neisseria gonorrhoeae* (NG) specific DNA.

The FlexStar® CT & NG PCR Detection Mix 1.5 is based on real-time PCR technology, utilizing polymerase chain reaction (PCR) for the amplification of CT and NG specific target sequences and fluorescently labeled target specific probes for the detection of the amplified DNA.

In addition to the CT and NG DNA specific amplification and detection systems, the FlexStar® CT & NG PCR Detection Mix 1.5 includes oligonucleotides for the amplification and detection of the internal control (IC, AltoStar® Internal Control 1.5). The IC is automatically added at the beginning of the nucleic acid purification procedure on the AltoStar® Automation System AM16 (in the following summarized as AltoStar® AM16). For details refer to the instructions for use of the AltoStar® Internal Control 1.5.

For CT, a dual-target system, targeting the chromosomal and plasmid DNA, is used. Both targets are detected in the same detection channel. For NG 2 targets are detected in 2 separate detection channels.

Probes specific for CT DNA are labeled with the fluorophore FAM™, probes specific for NG Target opa (NG opa) DNA are labeled with the fluorophore ROX™ and probes specific for NG Target porA (NG porA) DNA are labeled with the fluorophore Cy5, respectively. The probe specific for the IC is labeled with the fluorophore JOE™.

Using probes linked to distinguishable dyes enables the parallel detection of CT, NG and the IC in the corresponding detection channels of the real-time PCR instrument.

4.1 Components

The FlexStar® CT & NG PCR Detection Mix 1.5 contains enough reagents for 96 reactions. The product consists of the following components:

- Detection Mix¹⁾
- PC²⁾
- NTC³⁾

¹⁾ Contains biological material of animal origin

²⁾ Positive Control (CT and NG specific DNA)

³⁾ No Template Control (negative control)

Except for the DNA polymerase, which is included in the FlexStar® (RT-)PCR Amplification Mix 1.5, the Detection Mix component contains all reagents (PCR buffer, magnesium salt, primers and probes) to allow detection and differentiation of CT and NG specific DNA, as well as of IC specific DNA.

The PC contains CT, NG opa as well as NG porA specific DNA. It is used to verify the functionality of the CT and NG DNA specific amplification and detection systems.

The NTC contains neither CT nor NG specific DNA but does contain the IC template. The NTC is used as negative control for the CT and the NG DNA specific real-time PCR and indicates possible contamination of the Detection Mix component.

4.2 Real-time PCR instruments

The FlexStar® CT & NG PCR Detection Mix 1.5 can be used with the following real-time PCR instruments:

- CFX96™ Deep Well Dx System (Bio-Rad)
- CFX96™ Dx System (Bio-Rad)
- QuantStudio™ 5 Real-Time PCR System (Applied Biosystems)
- Rotor-Gene® Q5/6 plex Platform (QIAGEN)

NOTE



Ensure that all instruments used have been installed, calibrated, checked and maintained according to the manufacturer's instructions and recommendations.

5. Material required but not provided

The following additional instruments and consumables are required for use of the FlexStar® CT & NG PCR Detection Mix 1.5 but not provided with this product:

- Appropriate real-time PCR instrument (see chapter 4.2 Real-time PCR instruments)
- Appropriate nucleic acid extraction system or kit (see chapter 6.1 Sample preparation)
- Vortex mixer
- Centrifuge (e.g. desktop centrifuge) for centrifugation of kit reagents
- Centrifuge for centrifugation of PCR plates
- Appropriate 96 well reaction plates or reaction tubes with corresponding (optical) closing material
- Pipettes (adjustable)
- Pipette tips with filters (disposable)
- Powder-free gloves (disposable)

Reagents required but not included in the FlexStar® CT & NG PCR Detection Mix 1.5:

- FlexStar® (RT-)PCR Amplification Mix 1.5 (Order No. FS0011503/FS0011505)
- AltoStar® Internal Control 1.5 (Order No. IC15-06)

6. Procedure

6.1 Sample preparation

Extracted DNA is the starting material for the FlexStar® CT & NG PCR Detection Mix 1.5. The quality of the extracted DNA has a profound impact on the performance of the product.

For additional information and technical support regarding pre-treatment and sample preparation, contact Altona Diagnostics technical support (see chapter 9. Technical assistance).

6.2 Master mix setup

All reagents and samples should be thawed completely, mixed (by pipetting or gentle vortexing) and centrifuged briefly before use.

The FlexStar® CT & NG PCR Detection Mix 1.5 is configured for use with the FlexStar® (RT-)PCR Amplification Mix 1.5 and the AltoStar® Internal Control 1.5, which allows to control the sample preparation procedure (nucleic acid extraction) and the subsequent PCR.

- ▶ The IC is automatically added at the beginning of the nucleic acid purification procedure on the AltoStar® AM16.
- ▶ When using other nucleic acid extraction methods, the IC has to be added during the lysis step either manually or automatically by the respective instrument.

- ▶ No matter which method/system is used for nucleic acid extraction, never add the IC directly to the sample. The IC should always be added to the sample/lysis buffer mixture. The volume of the IC which has to be added, always and only depends on the elution volume. It represents 50 % of the elution volume. For instance, if the nucleic acid is going to be eluted in 60 µl of elution buffer or water, 30 µl of IC per sample must be added into the sample/lysis buffer mixture.
- ▶ Set up the master mix according to the following pipetting scheme:

Table 2: Pipetting scheme (master mix setup)

Number of reactions (rxns)	1	12
Detection Mix	5 µl	60 µl
Amplification Mix	15 µl	180 µl
Volume master mix	20 µl	240 µl

NOTE

No matter which method/system is used for nucleic acid extraction, never add the IC directly to the specimen.

6.3 Reaction setup

- ▶ Pipette 20 µl of the master mix into each required well of an appropriate optical 96-well reaction plate or an appropriate optical reaction tube.
- ▶ Add 10 µl of the sample (eluate from the nucleic acid extraction) or 10 µl of the controls (PC or NTC).

Table 3: Pipetting scheme (reaction setup)

Reaction setup	
Master mix	20 µl
Sample or control	10 µl
Total volume	30 µl

- ▶ Make sure that at least 1 PC and 1 NTC is used per run.
- ▶ Thoroughly mix the samples and controls with the master mix by pipetting up and down.
- ▶ Close the 96-well reaction plate with appropriate lids or optical adhesive film and the reaction tubes with appropriate lids.
- ▶ Centrifuge the 96-well reaction plate in a centrifuge with a microtiter plate rotor and the reaction tubes in an appropriate centrifuge for 30 seconds at approximately 1,000 x g (~ 3,000 rpm).
- ▶ The NTC does already contain the IC template in the correct concentration.

7. Programming the real-time PCR instrument

For basic information regarding the setup and programming of the different real-time PCR instruments, refer to the user manual of the respective instrument.

For detailed programming instructions regarding the use of the FlexStar® CT & NG PCR Detection Mix 1.5 on specific real-time PCR instruments, contact Altona Diagnostics technical support (see chapter 9. Technical assistance).

7.1 Settings

- ▶ Define the following settings:

Table 4: Run settings

Settings	
Reaction volume	30 µl
Ramp rate	Default
Passive reference*	None

* If applicable

7.2 Fluorescence detectors (dyes)

- ▶ Define the fluorescence detectors (dyes):

Table 5: Fluorescence detectors

Target	Detector name	Reporter	Quencher
CT specific DNA	CT	FAM™	(None)
NG opa specific DNA	NG opa	ROX™	(None)
NG porA specific DNA	NG porA	Cy5	(None)
Internal Control	Internal Control	JOE™	(None)

7.3 Temperature profile and dye acquisition

- Define the temperature profile and dye acquisition:

Table 6: Temperature profile and dye acquisition

	Stage	Cycle repeats	Acquisition	Temperature [°C]	Time [min:s]
Reverse transcription	Hold	1	-	52	05:00
Denaturation	Hold	1	-	95	00:05
Amplification	Cycling	45	-	95	00:05
			Yes	58	00:25

8. Data analysis

For basic information regarding data analysis on specific real-time PCR instruments, refer to the user manual of the respective instrument.

For detailed instructions regarding the analysis of the data generated with the FlexStar® CT & NG PCR Detection Mix 1.5 on different real-time PCR instruments, contact altona Diagnostics technical support (see chapter 9. Technical assistance).

8.1 Interpretation of results

8.1.1 Qualitative analysis

Table 7: Result interpretation

Detection channel				Result interpretation
FAM™ (CT)	ROX™ (NG opa)	Cy5 (NG porA)	JOE™ (IC)	
+	+	+	+/-*	CT, NG opa and NG porA specific DNA detected. ¹⁾
+	+	-	+/-*	CT and NG opa specific DNA detected. ²⁾
+	-	+	+/-*	CT and NG porA specific DNA detected. ²⁾
-	+	+	+/-*	NG opa and NG porA specific DNA detected. ¹⁾
+	-	-	+/-*	Only CT specific DNA detected.
-	+	-	+/-*	Only NG opa specific DNA detected. ²⁾
-	-	+	+/-*	Only NG porA specific DNA detected. ²⁾
-	-	-	+	Neither CT nor NG specific DNA detected. The sample does not contain detectable amounts of CT or NG specific DNA.
-	-	-	-	PCR inhibition or reagent failure. Repeat testing from original sample or collect and test a new sample.

* Detection of the IC in the JOE™ detection channel is not required for positive results in the FAM™ and/or the ROX™ and/or the Cy5 detection channel. A high CT and/or NG DNA load in the sample can lead to reduced or absent IC signals.

¹⁾ For a positive NG result detection of NG opa specific DNA in the ROX™ detection channel and NG porA specific DNA in the Cy5 detection channel is required.

²⁾ Results can be inconclusive due to either low NG DNA load in the sample or genetic exchanges between NG and other *Neisseria* species. Sample collection, extraction and PCR might be repeated to clarify the NG infection status.

Data analysis using the Rotor-Gene® Q5/6 plex Platform (QIAGEN):

Using the Rotor-Gene® Q5/6 plex Platform (QIAGEN) a slight crosstalk between the orange (ROX™) detection channel and the red (Cy5) detection channel might be observed. This can lead to the following signal phenotype.

CT DNA specific signals will show up only in the green (FAM™) detection channel, NG porA DNA specific signals will show up only in the red (Cy5) detection channel, internal control specific signals will show up only in the yellow (JOE™) detection channel. In contrast, NG opa DNA specific signals might not only show up in the orange (ROX™) detection channel, but also generate a weak crosstalk signal in the red (Cy5) detection channel (see figure 1).

The intensity of the crosstalk signal of NG opa in the red (Cy5) detection channel directly correlates with the respective signal intensity in the orange (ROX™) detection channel (see figure 1). Therefore, we highly recommend analyzing the signal of a potential NG porA positive sample in the red (Cy5) detection channel in direct comparison with the respective NG opa signal of the same sample and the NG positive control. If you have any questions regarding the data analysis on a Rotor-Gene® Q5/6 plex Platform (QIAGEN), contact altona Diagnostics technical support (see chapter 9. Technical assistance).

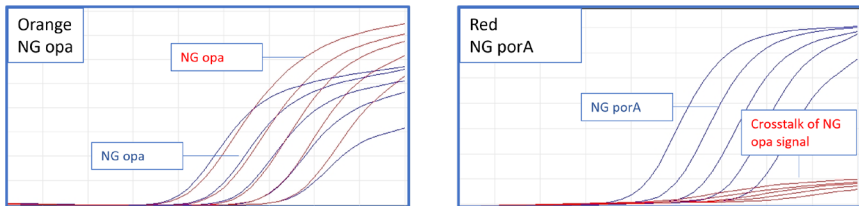


Figure 1: Dilution series of specific DNA containing target sequences of NG opa and NG porA (blue) or NG opa only (red) ranging from 1.00E+04 copies/μl to 1 copy/μl. The samples were analyzed using the FlexStar® CT & NG PCR Detection Mix 1.5 and the FlexStar® (RT-)PCR Amplification Mix 1.5 on a Rotor-Gene® Q5/6 plex Platform (QIAGEN). Samples of NG opa might generate signals not only in the orange (ROX™) detection channel, but also weak crosstalk signals in the red (Cy5) detection channel.

9. Technical assistance

For customer support, contact altona Diagnostics technical support:

e-mail: **support@altona-diagnostics.com**

phone: **+49-(0)40-5480676-0**

10. Trademarks and disclaimers













AltoStar®, FlexStar® (altona Diagnostics); QuantStudio™ (Applied Biosystems); CFX96™ (Bio-Rad); Rotor-Gene® (QIAGEN); FAM™, JOE™, ROX™ (Thermo Fisher Scientific).




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11. Explanation of symbols

Symbol	Explanation
	Research use only
	Batch code
	Content
	Cap color
	Catalogue number
	Number
	Component
	Consult instructions for use
	Contains sufficient for "n" tests/reactions (rxns)
	Temperature limit
	Use-by date
	Manufacturer

Symbol	Explanation
	Material number
	Version
	Note: Information is given to the user that is useful but not essential to the task at hand.

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